A Review: Antibiotic sensitivity of microbes obtained from various soil habitats

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Abstract

Soil is home to various microorganisms, including bacteria and fungi, which vary across different habitats. Research indicates that microbial populations exist in various layers of soil and are influenced by its physical and chemical properties. As the uppermost layer of the Earth's surface, soil supports numerous living organisms. Its chemical composition is shaped by factors such as geographical location, human activities, animal presence, and surrounding flora and fauna.

The extensive use of antibiotics in treating infectious diseases has led to their release into the environment through human and animal waste, improper disposal, and other sources. These antibiotics can accumulate in soil and water, potentially impacting microbial communities. Prolonged exposure to antibiotics may contribute to the development of resistance in soil-dwelling pathogens, altering their susceptibility to different antibiotics. As a result, various pathogens present in the soil may exhibit differing levels of resistance, raising concerns about the environmental impact of antibiotic contamination.

Keywords: Microorganisms, Antibiotics, Resistance, Pathogens, Contamination.

Introduction

The thin layer of material covering much of the Earth's surface is known as soil. Despite being fragile and often less than a meter thick, it plays a crucial role in sustaining life [1]. Recently, it has been described as "the most complicated biomaterial on the planet." This complexity arises from two key components: the abiotic soil structure and the biotic diversity, which together create physical and biological heterogeneity. Physically, soil is composed of sand, silt, and clay particles of varying sizes. The size differences, ranging from millimetre-sized sand particles to micrometre-sized clay particles, significantly impact the surface area of soil components, influencing soil chemistry, including chemical reactions and transformations. On the other hand, soil microflora regulates biochemical processes within the soil. Interestingly, the organic and biological components of soil resemble mineral components in terms of size. The primary soil biota, in increasing order of size, include viruses, bacteria, fungi, algae, and protozoa. As the size of organisms decreases, their numbers increase dramatically. Despite the vast number of microorganisms present in the soil, they cover only about 10⁻⁶% of the total surface area, a proportion comparable to the land area occupied by humans on Earth [1]. Soil microflora are essential for biochemical processes that support human life, such as plant growth, the production of bioactive compounds beneficial to human health, and groundwater protection [2-4]. However, soil is also a habitat for human pathogens and serves as a reservoir for bacterial antibiotic resistance. Bacterial

pathogens in soil are free-living organisms capable of reproducing in human or animal hosts. Some of the notable pathogens found in soil include *Campylobacter jejuni*, pathogenic strains of *Escherichia coli*, *Salmonella* species, *Shigella* species, *Vibrio cholerae*, and *Yersinia* species [5].

The desert soils of the Southwestern USA are distinct due to their arid environment. These soils typically have a high pH (>8.0) and may contain excessive salts and sodium. Such abiotic stress, along with the biotic stress exerted by native soil microorganisms, often results in the rapid die-off of introduced organisms, such as rhizobia, within a few weeks [6-9]. Furthermore, *E. coli* has been identified in tropical and subtropical soils. Laboratory studies have demonstrated that *E. coli* can grow in tropical soils under suitable conditions, particularly when provided with amendments [10-14].

Byappanahalli and colleagues reported that *E. coli* could also be isolated from coastal temperate forest soils in Indiana, indicating that soilborne *E. coli* may be more widespread than previously assumed [15]. Soil is increasingly subjected to environmental pressures that impact its ability to perform essential ecosystem services. To preserve these vital soil functions, it is crucial to understand how soil microorganisms respond to disturbances and environmental changes. The growing number of studies on soil microorganisms and their role in maintaining soil ecosystem stability is likely driven by the increasing focus on the relationship between biodiversity and ecosystem functioning. The loss of biodiversity has been recognized as a significant threat to soil (COM 231). However, due to the complexity and variability of soil, microbial ecologists face significant challenges in quantifying the role of microorganisms in soil resilience, and the mechanisms that enable soil to withstand disturbances remain poorly understood [16,17].

The extensive use of antibiotics in food animal production is considered a key factor contributing to the emergence of antibiotic-resistant bacteria (ARB) in the environment [18,19]. Studies estimate that up to 80% of orally administered antibiotics in animals may pass through their bodies unchanged, eventually accumulating in manure [20,21]. Reported antibiotic concentrations in manure range from trace levels to as high as 200 mg/L [22,23]. The persistence of antibiotics in soil is influenced by various factors, including soil type, climate, and the specific class of antibiotics. While most antibiotics degrade in soil, some have an extended half-life [24]. Antibiotics exert both quantitative and qualitative effects on native microbial communities in terrestrial environments [25]. Although the antibiotic concentrations in most soils are not high enough to have therapeutic effects, they may still contribute to the selection of antibiotic-resistant bacteria [25,26,27,28]. An increase in antibiotic resistance among soil bacteria following manure application may result from (i) the exchange of genetic elements between soil bacteria and antibiotic-resistant bacteria in manure, (ii) the transfer of genetic material among antibiotic-producing soil microorganisms, or (iii) selective pressure from low levels of antibiotics in manure. Many antibiotics also have a strong tendency to bind with soil [29,30]. In addition to beneficial components, sludge may contain indigenous populations of human enteric pathogens. Some of these enteric bacteria constitute a significant group of such pathogens. In a related study, it was demonstrated that the removal of water from sludge through evaporation to near dryness resulted in a reduction of most bacterial populations by approximately one order of magnitude or less [31].

Studies of microflora on Sludge

Pepper et al. conducted both laboratory and field studies to assess the survival and movement of bacterial pathogens introduced into soil through sludge application. Their findings indicate that factors such as soil moisture, texture, and temperature play a crucial role in determining the persistence of bacterial pathogens in sludge-amended soil [32].

A study assessed the abundance of bacteria and antibiotic resistance genes using both culture-dependent and molecular techniques. The findings suggest that growing vegetable crops in soil fertilized with human waste, without proper treatment or an adequate waiting period, may contribute to an increased presence of antibiotic resistance genes in harvested produce [33]. Some studies examined the impact of moisture content on the survival and regrowth of both seeded and indigenous enteric bacteria in raw sludge. The findings indicate that reducing sludge moisture leads to bacterial inactivation; however, in dried sludge (>90% solids), most bacteria remain stable except Proteus mirabilis. Indigenous bacteria can persist for extended periods in dried sludge. Bacterial regrowth occurs in sterilized sludge with \leq 75% solids but not in sludge with \geq 85% solids. Salmonella typhimurium can grow in both liquid and dewatered sludge, though at lower densities when indigenous bacteria are present, and it declines rapidly in their presence [34]. According to an assessment the impact of five annual liquid sewage sludge applications on the organic carbon and nitrogen content of a furrowirrigated desert soil. The findings suggest that liquid sludge application in desert soils may influence underground aquifers, potentially contributing to nitrate pollution [35]. A review by Timothy M. Straub examines pathogen types and concentrations in sludge, sludge treatment efficacy, pathogen fate after land disposal, exposure pathways, detection methods, risk assessment models, and future research needs. Findings indicate that despite stabilization and treatment, significant pathogens persist in sludge. If viable for extended periods, they may contaminate groundwater beneath disposal sites, travel through the vadose zone with minimal inactivation, and spread over significant distances [36].

Studies of microflora on antibiotic sensitivity and resistance

Brooks et al. investigated the prevalence of antibiotic-resistant bacteria and endotoxins in soil following biosolid application. The study established a baseline for assessing the impact of biosolid application on soil antibiotic-resistant bacteria and endotoxin levels [37]. Some study explored soil bacteria's ability to utilize antibiotics as a carbon source, their phylogenetic diversity, and their role as a reservoir for antibiotic resistance. Findings suggest that this overlooked reservoir may contribute to rising multidrug resistance in pathogens. The isolated bacteria exhibited extreme resistance, tolerating antibiotic concentrations over 50 times higher than standard levels [38]. Pepper et al.. examined the soil's role in public health, including its impact on pathogens, antibiotics, nutrition, and climate change. The study concludes that soil is a vital public health asset with an estimated value of \$20 trillion, making it the world's most valuable ecosystem [39]. Some authors assessed the impact of TWW irrigation on soil antibiotic-resistant (AR) bacteria and ARG reservoirs. Findings suggest that TWW-associated bacteria have a minimal effect on the soil microbiome, and high AR bacteria and ARG levels in both freshwater-and TWW-irrigated soils likely stem from native AR within the natural soil microbiome [40].

Fiona Walsh's study identified and characterized multidrug resistance (MDR) mechanisms in the culturable soil antibiotic resistome, linking resistance profiles to bacterial species. Findings highlight the key role of efflux mechanisms and differences in intrinsic resistance between clinical and soil bacteria of the same family [41]. Eddie Cytryn's study explored natural and human influences on soil antibiotic resistance and its transfer to pathogens, supporting the resistome hypothesis. The review offers a comprehensive overview of factors shaping soil antibiotic resistance [42]. Romain Marti's study assessed antibiotic-resistant bacteria on raw vegetables grown in soil fertilized with inorganic, dairy, or swine manure. Findings revealed viable coliform bacteria resistant to multiple antibiotics, even on vegetables from never-manured soil [43]. The study on human waste assessed bacterial abundance and antibiotic resistance genes using culture-dependent and molecular methods. Findings suggest that growing Vegetables in soil fertilized with untreated human waste increases antibiotic resistance genes in harvested crops [44]. Few studies explored strategies to discover novel antimicrobials from actinomycetes by activating biosynthetic gene clusters through genetic, chemical, and ecological methods. Findings highlight that many antibiotic-producing genes remain unexpressed under standard conditions, but genome sequencing reveals a vast untapped reservoir. Chemical elicitors, optimized culturing, and ecological approaches can help unlock these genes for new drug discovery [45].

A study was conducted on native Nebraska prairie soils unaffected by human or animal waste to assess background antibiotic resistance. Findings showed no correlation between resistance and soil physical or chemical properties [46]. A comparative study on the diversity, abundance, and composition of antibiotic resistance genes (ARGs) and bacteria in 12 urban parks in Victoria, Australia, with and without reclaimed water irrigation (RWI). Findings indicate that RWI significantly increased ARG abundance and diversity but did not notably enhance horizontal gene transfer potential [47].

An investigation examined the correlation between metal concentrations and antibiotic resistance genes. Findings show that even low levels of metals like aluminum, copper, manganese, and lead in residential soils promote antibiotic resistance, evidenced by increased gene abundances [48]. A review re-evaluates the impact of land application on antibiotic-resistant bacteria (ARB) and genes (ARG), highlighting their public health risks from environmental exposure. Findings suggest that while waste application temporarily increases resistance in soil, its persistence varies by site and is often inconsistent [49]. Another review examines antibiotic degradation in soil and its impact on microbial communities. Findings show that antibiotics alter enzyme activity, carbon metabolism, microbial biomass, and the balance of Gram-negative bacteria, Gram-positive bacteria, and fungi [50].

The impact of PFD on antibiotic resistance genes (ARGs) and bacterial communities in a lab-scale soil experiment was studied by reserchers. Redundancy analysis revealed that penicillin-induced shifts in the bacterial community primarily drive ARG composition [51]. A study analyzed soil bacterial composition and function in response to multidrug-resistant *E. coli* with and without tetracycline contamination. Findings indicate that antibiotic contamination worsens the impact of foreign ARB, posing risks to soil quality [52].

According to a study the activity of soil-bound tetracycline and tylosin on bacterial cultures in different soil types. Results show that despite strong adsorption to clay, antibiotics remain biologically active and may contribute to antibiotic resistance in soil [53]. An investigation improves the agar diffusion assay by incorporating antibiotic loss during diffusion, leading to more accurate MIC determination. The model is broadly applicable to other dissipative processes, such as antigen diffusion and substrate load calculations in affinity purification [54].

Microbial Diversity of soil

Ishii et al. found viable E. coli in northern temperate soils across three Lake Superior watersheds, with seasonal population variations. This study is the first to report naturalized E. coli growth in nonsterile, nonamended soils, challenging its reliability as a fecal contamination indicator [55].

An investigator examined the transfer and persistence of E. coli O157:H7 in lettuce grown in soil fertilized with contaminated compost or irrigated with contaminated water. Results showed that both sources could contaminate lettuce and soil, posing a health risk. Proper management of irrigation water, compost, and land history is crucial to preventing pathogen introduction [56]. A study explored factors influencing soil microbial resistance and resilience to environmental disturbances. Results indicate that soil stability depends on physicochemical structure, microbial composition, and biotic-abiotic interactions, offering a measurable indicator of soil health [57]. It was assessed in an experiment that the survival of three E. coli O157:H7 strains in four Chinese soils. Results showed that survival times ranged from 8.23 to 62.33 days, with biodiversity limiting E. coli invasion and virulence genes reducing survival [58].

Soil pathogens and human health

Ian L. Pepper explores the soil-health-human-health nexus, highlighting how soil influences human well-being and how human activities impact soil health, with strategies for improvement [59].

Studies regarding Physical factors of soil pressure

Zeyou Chen examined the bioavailability of tetracycline sorbed on three soil types to a fluorescent *E. coli* bioreporter, finding that soil-sorbed tetracycline can still exert selective pressure on bacteria, influenced by soil texture and water potential [60].

Conclusion

Soil, a highly complex and dynamic biomaterial, plays a fundamental role in sustaining life by supporting plant growth, regulating biochemical processes, and serving as a reservoir for both beneficial and harmful microorganisms. While soil microbiota contributes to essential ecosystem functions, it also harbors human pathogens such as *E. coli, Salmonella*, and *Campylobacter*, which can persist across diverse soil types. Additionally, environmental disturbances, particularly the widespread use of antibiotics in agriculture, have accelerated the emergence of antibiotic-resistant bacteria (ARB) in soil. Antibiotics excreted by livestock accumulate in manure and, when applied to soil, exert selective pressure on microbial communities, fostering the spread of resistance genes. Although most antibiotics degrade over time, their residual presence can alter microbial composition and resistance dynamics. Furthermore, land application of biosolids and manure may introduce enteric pathogens, raising concerns about potential human health risks. Given the critical role of soil in maintaining ecological balance, it is imperative to understand microbial interactions, enhance soil resilience, and adopt sustainable management practices to mitigate the spread of antibiotic resistance while preserving essential soil functions.

References :

1. Young, J.M., and Crawford, J.W. (2004). Interactions and self-organization in the soil-microbe complex. Science, 304, 1634.

2. Stirzaker, R.J., Passioura, J.B., and Wilms, Y. (1996). Soil structure and plant growth: Impact of bulk density and biopores. Plant Soil, 185, 151

3. Strobel, G., and Daisy, B. (2003). Bioprospecting for microbial endophytes and their natural products. Microbiol. Molec. Biol. Rev., 67, 491.

4. Bejat, L., Perfect, E., Quisenberry, F.L., Coyne, M.S., and Haszler, G.R. (2000). Solute transport as related to soil structure in unsaturated intact soil blocks. Soil Sci. Soc. Am. J., 64, 818.

5. Ward RL, McFeters GA, and Yeager JG. Pathogens in Sludge: Occurrence, Inactivation, and Potential for Regrowth. United States Department of Energy. 1984; DE-AC 04-76 DPOO789.

6. Pillai SD and Pepper IL. Survival of Tn5 Mutant Bean Rhizobia in Desert Soils: Phenotypic Expression of Tn5 under Moisture Stress. Soil Biol. Biochem. 1990; 22:265-270.

7. Pillai SD and Pepper IL. Transposon Tn5 as an Identifiable Marker in Rhizobia: Survival and Genetic Stability of Tn5 Mutant Bean Rhizobia under Temperature Stressed Conditions in Desert Soils. Microb. Ecol. 1991; 21:21-33.

8. Miller MS, and Pepper IL. Survival of a Fast Growing Strain of Lupine Rhizobia in Sonoran Desert Soil. Soil Biol. Biochem. 1988; 20:319-322.

9. Shoushtari, NH, and Pepper IL. Mesquite Rhizobia Isolated from the Sonoran Desert. II. Competitiveness and Survival in Desert Soils. Soil Biol. Biochem. 1985; 17:803-806.

10. Byappanahalli, M. N., and R. S. Fujioka. 1998. Evidence that tropical soil environment can support the growth of Escherichia coli. Water Sci. Technol. 38:171–174

11. Desmarais, T. R., H. M. Solo-Gabriele, and C. J. Palmer. 2002. Influence of soil on fecal indicator organisms in a tidally influenced subtropical environment. Appl. Environ. Microbiol. 68:1165–1172.

12. Fujioka, R. S. 2001. Monitoring coastal marine waters for spore-forming bacteria of faecal and soil origin to determine point from non-point source pollution. Water Sci. Technol. 44:181–188.

13. Solo-Gabriele, H. M., M. A. Wolfert, T. R. Desmarais, and C. J. Palmer. 2000. Sources of Escherichia coli in a coastal subtropical environment. Appl. Environ. Microbiol. 66:230–237

14. Byappanahalli, M., and R. Fujioka. 2004. Indigenous soil bacteria and low moisture may limit but allow faecal bacteria to multiply and become a minor population in tropical soils. Water Sci. Technol. 50:27–32.

15. Byappanahalli, M. N., R. L. Whitman, D. A. Shively, M. J. Sadowsky, and S. Ishii. 2006. Population structure, persistence, and seasonality of autochthonous Escherichia coli in temperate, coastal forest soil from a Great Lakes watershed. Environ. Microbiol. 8 [Online.] doi:10.1111/j.i4622920.2005.00916.x.

16. COM (2006)231. Communication from the Commission to the Council, the European Parliament, the European Economic and Social Committee and the Committee of Regions – Thematic Strategy for Soil Protection. Commission of the European Communities, Brussels.

17. Bryan S. Griffiths1 & Laurent Philippot2(2012). Insights into the resistance and resilience of the soil microbial community. FEMS (Federation of European Microbiological Societies) Microbiol Rev1–18.

18. Jørgensen, S.E., and B. Halling-Sørensen. 2000. Editorial, "Drugs in the environment Chemosphere 40:691–699.

19. Rooklidge, S.J. 2004. Environmental antimicrobial contamination from tetraccumulation and diffuse pollution pathways. Sci. Total Envi ron. 35:1–13.a

20. Levy, S.B. 1992. The antibiotic paradox: How miracle drugs are destroying the miracle. Plenum Publishing.

21. Thiele-Bruhn, S. 2003. Pharmaceutical antibiotic compounds in soils— A review. J. Plant Nutr. Soil Sci. 166:145–167.

22. Kumar, K., A. Thompson, A.K. Singh, Y. Chander, and S.C. Gupta. 2004. Enzyme linked immunosorbent assay for ultratrace determination of antibiotics in aqueous samples. J. Environ. Qual. 33:250–256.

23. Kumar, K.,S.C.Gupta, Y.Chander, and A.K.Singh. 2005. Antibiotics in agriculture and their impact on the terrestrial environment. Adv. Agron. 87 (in press).

24. Boxall, A.B., L.A. Fogg, P.A. Blackwell, P. Kay, E.J. Pemberton, and A. Croxford. 2004. Veterinary medicines in the environment. Rev. Environ. Contam. Toxicol. 180:1–91.

25. Nygaard, K., B.T. Lunestad, H.Hektoen, J.A.Berge, and V. Hormaza bal. 1992. Resistance to oxytetracycline, oxolinic acid, and furazoli done in bacteria from marine sediments. Aquaculture 104:31–36.

26. USEPA.2002.Environmental and Economic Benefit Analysis of Final Revisions to the National Pollutant Discharge Elimination System Regulation and the Effluent Guidelines for Concentrated Animal Feeding Operations. EPA 821-R-03-003. USEPA Office of Water, Washington, DC.

27. Gavalchin, J., and S.E. Katz. 1994. The persistence of fecal-borne antibiotics in soil. J. AOAC Int. 77:481–485.

28. Kummerer, K. 2003. Significance of antibiotics in the environment. J. Antimicrob. Chemother. 52:5–7.

29. Tolls, J. 2001. Sorption of veterinary pharmaceuticals in soil: A review. Environ. Sci. Technol. 35:3397–3406.

30. Kumar,K., S.C.Gupta, Y.Chander, and A.K.Singh.2005.Antibiotics in agriculture and their impact on the terrestrial environment. Adv. Agron. 87 (in press).

31. Ward, R. L, J. G. Yeager, and C. S. Ashley. 1981. Response of bacteria in wastewater sludge to moisture loss by evaporation and effect of moisture content on bacterial inactivation by ionizing radiation. Apl. Envi. ron. Microbiol. 41:1123-1127.

32. Pepper I.L., Karen L. Josephson, Rachel L. Bailey, Mark D. Burr and Charles P. Gerba (1993) SURVIVAL OF INDICATOR ORGANISMS IN SONORAN DESERT SOIL AMENDED WITH SEWAGE SLUDGE, Journal of Environmental Science and Health. Part A: Environmental Science and Engineering and Toxicology: Toxic/ Hazardous Substances and Environmental Engineering, vol. A28(6), p.1287-1302.

33. Teddie O. Rahube, Romain Marti, Andrew Scott, Yuan-Ching Tien, Roger Murray, Lyne Sabourin, Yun Zhang, Peter Duenk, David R. Lapen, Edward Toppa, (2014) Impact of Fertilizing with Raw or Anaerobically Digested Sewage Sludge on the Abundance of Antibiotic-Resistant Coliforms, Antibiotic Resistance Genes, and Pathogenic Bacteria in Soil and on Vegetables at Harvest, Applied and Environmental Microbiology Volume 80 Number 22 p. 6898 – 6907

34. J. GARY YEAGER AND RICHARD L. WARD (1981) Effects of Moisture Content on Long-Term Survival and Regrowth of Bacteria in Wastewater Sludge, APPLED AND ENVIRONMENTAL MICROBIOLOGY, Vol. 41, No. 5, p. 1117-1122

35. J.E. Artiola and I.L. Pepper (1992) Long-term influence of liquid sewage sludge on the organic carbon and nitrogen content of a furrow-irrigated desert soil Biology and Fertility of Soils 14:30-36.

36. Timothy M. Straub, t Ian L. Pepper, and Charles P. (1993) Gerba Hazards from Pathogenic Microorganisms in Land-Disposed Sewage Sludge, Reviews of Environmental Contamination and Toxicology, Vol. 132.

37. J.P. Brooks, S.L. Maxwell, C. Rensing, C.P. Gerba, and I.L. Pepper (2007) Occurrence of antibiotic-resistant bacteria and endotoxin associated with the land application of biosolids.Can. J. Microbiol. 53: 616–622,doi:10.1139/W07-021.

38. Gautam Dantas, Morten O. A. Sommer, Rantimi D. Oluwasegun,1 George M. Church1(2008) Bacteria Subsisting on Antibiotics, VOL 320 SCIENCE <u>www.sciencemag.org</u>

39. L. PEPPER, C. P. GERBA, D. T. NEWBY, and C. W. RICE, (2009) Soil: A Public Health Threat or Savior Critical Reviews in Environmental Science and Technology, Vol. 39: p.416–432.

40. Yael Negreanu, Zohar Pasternak, Edouard Jurkevitch, and Eddie Cytryn, (2012)Impact of Treated Wastewater Irrigation on Antibiotic Resistance in Agricultural Soils, Environmental Science & Technology, Vol. 46, p. 4800–4808 dx.doi.org/10.1021/es204665b

41. Fiona Walsh, Brion Duffy (2013) The Culturable Soil Antibiotic Resistome: A Community of Multi-Drug Resistant Bacteria, PLOS ONE | www.plosone.org | Volume 8 | Issue 6 | e65567

42. Eddie Cytryn, The soil resistome: The anthropogenic, the native, and the unknown (2013) Soil Biology & Biochemistry Vol. 63 p 18-23

43. Romain Marti, Andrew Scott, Yuan-Ching Tien, Roger Murray, Lyne Sabourin, Yun Zhang, Edward Topp (2013) Applied and Environmental Microbiology, Vol 79 Number 18 p. 5701–5709

44. Teddie O. Rahube, Romain Marti, Andrew Scott, Yuan-Ching Tien, Roger Murray, Lyne Sabourin, Yun Zhang, Peter Duenk, David R. Lapen, Edward Toppa, (2014) Impact of Fertilizing with Raw or Anaerobically Digested Sewage Sludge on the Abundance of Antibiotic-Resistant Coliforms, Antibiotic Resistance Genes, and Pathogenic Bacteria in Soil and on Vegetables at Harvest, Applied and Environmental Microbiology Volume 80 Number 22 p. 6898 – 6907

45. Hua Zhu • Stephanie K. Sandiford • Gilles P. van Wezel (2014) Triggers and cues that activate antibiotic production by actinomycetes, J Ind Microbiol Biotechnology 41:371–386

46. Lisa M. Durso, David A. Wedin, John E. Gilley, Daniel N. Miller, and David B. Marx (2015) Assessment of Selected Antibiotic Resistances in Ungrazed Native Nebraska Prairie Soils, Journal of Environmental Quality, doi:10.2134/jeq2015.06.0280

47. Xue-Mei Han, Hang-Wei Hu, Xiu-Zhen Shi, Jun-Tao Wang, Li-Li Han, Deli Chen, Ji-Zheng He (2016) Impacts of reclaimed water irrigation on soil antibiotic resistome in urban parks of Victoria, Australia, Environmental Pollution Vol. 211 p. 48-57 http://dx.doi.org/10.1016/j.envpol.2015.12.033 0269-7491/© 2015 Elsevier Ltd. All rights reserved

48. Charles W Knapp & Anna C Callan & Beatrice Aitken & Rylan Shearn & Annette Koenders & Andrea Hinwood (2017) Relationship between antibiotic resistance genes and metals in residential soil samples from Western Australia, Environ Sci Pollut Res 24:2484–2494 DOI 10.1007/s11356-016-7997-y

49. Ian Pepper, John P. Brooks, and Charles P. Gerba (2018) Antibiotic resistant bacteria in municipal wastes: Is there reason for concern? Environ. Sci. Technol., Vol 52/Issue 7 p. 1-46

50. Mariusz Cycon', Agnieszka Mrozik and Zofia Piotrowska-Seget (2019) Antibiotics in the Soil Environment— Degradation and Their Impact on Microbial Activity and Diversity, Frontiers in Microbiology, Sec. Microbiotechnology Volume 10 Article 338 <u>https://doi.org/10.3389/fmicb.2019.00338</u>

51. Bing Wang, Jianquan Yan, Guomin Li, Jian Zhang, Lanhe Zhang, Zheng Li, Houhe Chen (2020) Risk of penicillin fermentation dreg: Increase of antibiotic resistance genes after soil discharge, Environmental Pollution 259, 113956 https://doi.org/10.1016/j.envpol.2020.113956 0269-7491/© 2020 Elsevier Ltd. All rights reserved.

52. Han Xu, Zeyou Chen, Xinyan Wu, Lin Zhao, Nan Wang, Daqing Mao, Hongqiang Ren, Yi Luo (2021) Antibiotic contamination amplifies the impact of foreign antibiotic-resistant bacteria on soil bacterial community, Science of the Total Environment Vol 758, https://doi.org/10.1016/j.scitotenv.2020.143693.

53. Yogesh Chander, Kuldip Kumar, Sagar M. Goyal, and Satish C. Gupta (2005) Antibacterial Activity of Soil-Bound Antibiotics, J. ENVIRON. QUAL., VOL. 34:1952–1957, doi:10.2134/jeq2005.0017

54. Boyan Bonev, James Hooper and Judicae Parisot (2008) Principles of assessing bacterial susceptibility to antibiotics using the agar diffusion method, Journal of Antimicrobial Chemotherapy 61, 1295–1301 doi:10.1093/jac/dkn090

55. Satoshi Ishii, Winfried B. Ksoll, Randall E. Hicks, and Michael J. Sadowsky (2006) Presence and Growth of Naturalized Escherichia coli in Temperate Soils from Lake Superior Watersheds, APPLIED AND ENVIRONMENTAL MICROBIOLOGY, p. 612–621 Vol. 72, No. 1.

56. M.Oliveira, I.Viñas, J.Usall, M. Anguera, M. Abadias (2012) Presence and survival of Escherichia coli O157:H7 on lettuce leaves and in soil treated with contaminated compost and irrigation water, International Journal of Food Microbiology Vol. 156 p. 133–140.

57. Bryan S. Griffiths & Laurent Philippot (2013) Insights into the resistance and resilience of the soil microbial community, Federation of European Microbiological Societies, MICROBIOLOGY REVIEWS 1–18, DOI: 10.1111/j.1574-6976.2012.00343.x

58. Jiajia Xinga, Haizhen Wanga, Philip Brookesa, Joana FalcãoSallesc, JianmingXu (2018) Soil pH and microbial diversity constrain the survival of E.coli in soil <u>Soil Biology and Biochemistry</u> <u>Volume 128</u>, Pages 139-149 https://doi.org/10.1016/j.soilbio.2018.10.013

59. IAN L. PEPPER, The Soil Health-Human Health Nexus(2013) Critical Reviews in Environmental Science and Technology, 43:2617–2652 DOI:10.1080/10643389.2012.694330

60. Zeyou Chen, Wei Zhang, Gang Wang, Yingjie Zhang, Yanzheng Gao, Stephen A. Boyd, Brian J. Teppen, James M. Tiedje, Dongqiang Zhu, and Hui Li, (2017) Bioavailability of Soil-Sorbed Tetracycline to Escherichia coli under Unsaturated Conditions, Environmental Science & Technology Vol 51/ Issue 11 http://dx.doi.org/10.1021/acs.est.7b00590