NOVEL STRESS INDICATING RP-HPLC METHOD FOR SIMULTANEOUS ESTIMATION OF FINASTERIDE AND TADALAFIL

Dr Vamseekrishna Gorijavolu* Salman khan SV Kali Dinesh , Paidipamula Tarun Kumar Department of Pharmaceutical Analysis,NRI College of Pharmacy,Pothavarappadu(V),Agiripalli (M),Eluru(D.t), Andhra Pradesh, India-521212

Abstract: A novel stability indicating RP-HPLC method was developed for the simultaneous estimation of Finasteride (FSD) and Tadalafil (TDF) in pharmaceutical dosage form. Effective chromatographic separation achieved using a Waters Luna Phenyl Hexyl C18 column (250 mm x 4.6 internal diameter, 5μ particle size) with mobile phase Acetonitrile: Ammonium formate pH-3.0 with OPA (70:30 % v/v) at a flow rate of 1 ml/min. The analytes were monitored at 228 nm using a photo diode array detector at ambient temperature. The retention times for FSD and TDF were about 2.415 and 7.263 mins, respectively. Calibration curves were linear in the ranges of 12.5 to 75 μ g/ml for FSD and 12.5 to 75 μ g/ml for TDF with correlation coefficients > 0.99. The method has the requisite accuracy, selectivity, sensitivity, precision and robustness .Degradation products obtained from the stress studies did not interfere with the detection of FSD and TDF. The validated RP-HPLC method was effectively used to the analysis of FSD and TDF in marketed dosage form. **Key words:** Finasteride, Tadalafil, RP-HPLC, Stability indicating

INTRODUCTION

Finasteride (FSD), Finasteride, also known as 17β -(N-tert-butylcarbamoyl)-4-aza- 5α -androst-1-en-3-one, is a synthetic androstane steroid and 4-azasteroid. It is an analogue of androgen steroid hormones like testosterone and DHT. **Fig. 1a**, is a specific inhibitor of steroid Type II 5α reductase, an intracellular enzyme that converts the androgen testosterone into 5α dihydrotestosterone. It is commonly used to treat benign prostatic hyperplasia, prostate cancer, and androgenetic alopecia..

Tadalafil (TDF), chemically (6R,12aR)-6-(1,3-Benzodioxol-5-yl)-2-methyl-2,3,6,7,12,12ahexahydropyrazino[1',2':1,6]pyrido[3,4-b]indole-1,4-dione, **Fig.1b** is a potent and selective phosphodiesterase-5 (PDE-5) inhibitor, a secondary messenger for the smooth muscle-relaxing effects of nitric oxide, which plays an important role in the vasodilation of erectile tissues. There are different methods like UV, Liquid chromatography-tandem mass spectrometric (LC–MS/MS) methods were developed for the simultaneous determination of FSD and TDF.LC–MS/MS is a specific instrumental based technique affects the cost and the speed of the quantitative analysis of the compounds of interest. Therefore, authors are made an attempt to develop the new stability indicating RP-HPC method for simultaneous determination of FSD and TDF in pure and pharmaceutical formulation.



Figure: 1a Finasteride



Figure 1b: Tadalafil

Materials and Methods

EXPERIMENTAL

FSD was procured from Dr Reddy's Laboratories and TDF was supplied by Rakshith Pharmaceuticals, Hyderabad as gift samples. All the reagents used were of HPLC grade.

Equipment

Analysis was carried out by using Waters Alliance HPLC 2695 System fitted with quaternary pumps, Photo Diode Array Detector, and Auto Sampler integrated with Empower 2 software.

Chromatographic Conditions

Mobile phase consisting of Acetonitrile: Ammonium formate pH-3.0 with OPA (70:30 % v/v) and it was filtered through nylon disc filter of $0.45\mu m$ (Millipore) and sonicated for 3 min before use. The flow rate was 1 mL/min and the injection volume was $10\mu L$. PDA detection was performed at 228 nm and the separation was achieved at ambient temperature.

Pharmaceutical formulations

A commercial product (EntadfiTM labeled contains 5 mg FSD and 5 mg TDF per capsule) was studied

Preparation of standard stock solution

Accurately weighed quantities (5mg each) of FSD and TDF were dissolved separately in sufficient quantity of mobile phase in a 10mL volumetric flask. The volume was adjusted up to the mark with mobile phase to obtain a stock solution of mg/mL each of FSD and TDF.

RESULTS AND DISCUSSION

Method Development

Several HPLC - UV analytical methods were published for the estimation of FSD and TDF alone or in combination with other drugs in bulk and pharmaceutical dosage forms and there were no methods reported on FSD and TDF combination. Also the published methods were not economical. The aim of the present work was to develop and validate a simple, efficient, sensitive and selective method for the simultaneous estimation of FSD and TDF in bulk and dosage form. In the present investigation, initial trials were made to develop LC conditions for the separation of FSD and TDF using Acetonitrile and 0.1% OPA (80:20 v/v) at a flow rate of 1.0 mL/min, With X-Bridge Phenyl (150x4.6 mm,3.5µ) at a flow rate of 1.0 mL/min and the Peaks are not separated properly. In another trial using Acetonitrile : Ammonium formate pH-3.0 with OPA (60:40 v/v) at a flow rate 1.0 ml/min, with Luna Phenyl Hexyl (250x4.6 mm, 5µ) column Plate count is not within the limit. Whereas, with the same column with change in mobile phase to Acetonitrile: Ammonium formate pH-3.0 with OPA (65:35 v/v) at a flow rate of 1.0 mL/min FSD was eluted at 2.291 min and TDF was at 8.719 min with broad peak shape. Then by changing mobile phase composition of (70:30 v/v), a good resolution with sharp peaks were obtained (Figure 2), and these conditions were finalized for simultaneous estimation of FSD and TDF in bulk and pharmaceutical dosage forms. For quantitative analytical purpose wavelength was set at 228 nm, which provided better reproducibility with minimum or no interference.

Method validation

The method described above has been validated as per the ICH guidelines (ICH–Guidelines Q2B, Switzerland, 1996) and the results were summarized below.

Specificity

Specificity of an analytical method is ability to measure specifically the analyte of interest without interference from blank and known impurities. For this purpose blank chromatogram, standard chromatogram and sample chromatogram were recorded. The chromatogram of blank shows no response at the retention times of drugs which confirms the response of drugs was specific.



System suitability

System suitability was carried out by injecting 50 μ g/mL of FSD and TDF at different injection volumes in the range of 12.5 μ g/mL to 75 μ g/mL. With increment of injection volumes, the %RSD for tailing factor and theoretical plate number was less than 1% and is satisfactory and data is given in **Table 1**.

| S.No | Parameter | Finasteride | Tadalafil | Accepted limit |
|------|--------------------|-------------|-----------|------------------------|
| 1 | Capacity factor(K) | 3.67 | 2.64 | <mark>></mark> 2 |
| 2 | Plate count(N) | 5349 | 5876 | <mark>>2000</mark> |
| 3 | Tailing factor(Tf) | 1.15 | 1.04 | <mark>< 2.0</mark> |
| 4 | Resolution(Rs) | | 21.50 | <mark>>2.0</mark> |
| 5 | %RSD | 0.52 | 0.48 | <mark>< 2.0%</mark> |

Table 1: System suitability parameters for Finasteride & Tadalafil



Linearity

A linear relationship was evaluated across the range (12.5-75 μ g/mL) of the analytical procedure in triplicate. The range of concentrations was selected based on 50-150 % of the test concentration (for assay). Peak area and concentrations were subjected to least square regression analysis to calculate regression equation. The regression coefficient (R²) for both the drugs were found to be > 0.99 and indicating good linearity. The linearity data is given in **Table 2** and **Figure 2&3**.

Table 2: Linearity of Finasteride and Tadalafil

| S.No | Finasteride | | Tadalafil | | |
|------|--------------|------------------|--------------|-----------|--|
| | Conc.(µg/ml) | Pea k area | Conc.(µg/ml) | Peak area | |
| 1 | 12.50 | 588487 | 12.50 | 618837 | |
| 2 | 25.00 | 1161250 | 25.00 | 1222790 | |
| 3 | 37.50 | 1751664 | 37.50 | 1718985 | |
| 4 | 50.00 | 2330781 | 50.00 | 2361081 | |

| 5 | 62.50 | 2863848 | 62.50 | 2952775 | |
|-----------------------|-------------------|---------|---------------------|---------|--|
| 6 | 75.00 | 3519293 | 75.00 | 3504261 | |
| Regressin equation | y = 46494 x +1793 | | y = 46307 x + 37204 | | |
| Slope | 46494 | | 46307 | | |
| Intercept | 1793 | | 37204 | | |
| R ² | 0.999 | | 0.999 | | |



Fig 2: Linearity curve of Finasteride



Fig 3 : Linearity curve of Tadalafil

Precision

Precision studies were carried out in terms of repeatability and reproducibility. Six replicate determinations were carried out and percent relative standard deviation for both the drugs was less than 2%, indicating the high degree of precision and results were given in **Table 3**.

| S. No. | Area for Finasteride | Area for Tadalafil |
|-----------------------|----------------------|--------------------|
| 1 | 2359715 | 2341128 |
| 2 | 2380789 | 2352087 |
| 3 | 2369223 | 2344278 |
| 4 | 2345136 | 2365687 |
| 5 | 2389054 | 2311713 |
| 6 | 2351732 | 2332174 |
| Average | 2365942 | 2341178 |
| Standard Deviation | 16964.39 | 18322.49 |
| %RSD | 0.72 | 0.78 |

Table 3: Method Precision for Finasteride & Tadalafil

Accuracy (Recovery)

Accuracy was determined by the recovery study of known amounts of FSD and TDF standards added to a sample solution. Different concentrations of the two active ingredients were added to the sample and the recovery was measured. The data obtained for the evaluation of linearity were used. The accuracy as reflected from the recovery data and statistical evaluation of the assay for the two active ingredients is listed in Table 4&5.

Table 4:Accuracy results of Finasteride

| %Concentration (at specification Level) | Area | Amount Added (mg) | Amount Found (mg) | % Recovery | Mean Recovery |
|---|---------|-------------------------|-------------------------|---------------|------------------|
| 50% | 1183115 | 2.5 | 2.5 | 100.0 | |
| 100% | 2355687 | 5 | 4.98 | 99.6 | 99.57 |
| 150% | 3515124 | 7.5 | 7.43 | 99.1 | |

 Table 5:The Accuracy results for Tadalafil

| %Concentratio n (at specification Level) | Area | Amount Added (mg) | Amount Found (mg) | % Recovery | Mean Recovery |
|---|---------|-------------------------|-------------------------|---------------|------------------|
| 50% | 1180354 | 2.5 | 2.53 | 101.2 | |
| 100% | 2328120 | 5 | 5 | 100.0 | 100.1 |
| 150% | 3468324 | 7.5 | 7.44 | 99.2 | |

LOD and LOQ

LOD and LOQ were calculated from the average slope and standard deviation of y-intercepts of the calibration curve. LOD was found to be 0.143 μ g/mL and 0.157 μ g/mL respectively for FSD and TDF, LOQ was found to be 0.472 μ g/mL and 0.534 μ g/mL respectively for FSD and TDF and data is given in **Table 6**.

| rable of Sensitivity parameters (LOD & LOQ) | | | | | | | |
|---|------------|-----------|--|--|--|--|--|
| Name of | LOD(µg/ml) | LOQ(µg/ml | | | | | |
| drug | |) | | | | | |
| Finasteride | 0.143 | 0.157 | | | | | |

 Table 6: Sensitivity parameters (LOD & LOQ)
 Code

| Tadalafil | 0.472 | 0.534 |
|-----------|-------|-------|
|-----------|-------|-------|

Robustness

Robustness of the analytical method is the indication of its reliability. The method has the capacity to remain unaffected with deliberate changes as there was no signicant change in the peak area with the change in flow rate and organic phase of mobile phase. There was slight change in retention time with the change in flow rate and the other chromatographic parameters were not altered. The data is given in **Table 7&8**.

| | Finasteride | | | | | | | |
|-----------------|----------------------|------------------------|-----------|------------|-------------|----------------|--|--|
| Parameter | Condition | Retention time(min) | Peak area | Resolution | Tailin g | Plate count | | |
| Flow rate | Less flow (0.8ml) | 2.684 | 2553564 | | 1.19 | 5444 | | |
| Change | Actual (1ml) | 2.415 | 2359707 | | 1.17 | 5342 | | |
| (mL/min) | More flow (1.2ml) | 2.290 | 2194138 | | 1.12 | 5279 | | |
| Organia | Less Org (63:37) | 2.700 | 2756501 | | 1.16 | 5470 | | |
| Phase change | Actual (70:30) | 2.418 | 2380724 | | 1.15 | 5355 | | |
| | More Org (77:23) | 2.225 | 1965248 | | 1.08 | 5247 | | |

Table7:Robustness results of Finasteride by RP-HPLC

| Descourse | Tadalafil | | | | | | | |
|-------------------------|----------------------|------------------------|-----------|------------|---------|----------------|--|--|
| Parameter | Condition | Retention time(min) | Peak area | Resolution | Tailing | Plate count | | |
| Flow rate | Less flow (0.8ml) | 7.685 | 2544562 | 23.01 | 1.13 | 5972 | | |
| Change | Actual (1ml) | 7.263 | 2325171 | 21.53 | 1.09 | 5869 | | |
| (mL/min) | More flow (1.2ml) | 6.958 | 2073124 | 22.68 | 1.06 | 5808 | | |
| | Less Org (63:37) | 7.711 | 2627541 | 23.18 | 1.14 | 5942 | | |
| Organic Phase change | Actual (70:30) | 7.269 | 2339721 | 21.50 | 1.10 | 5862 | | |
| | More Org (77:23) | 6.606 | 1822874 | 21.30 | 1.04 | 5776 | | |

| Table 8: | Robustness | results | of | Tadalafil | bv | RP- | HPL | Æ |
|----------|----------------------|----------|-----|-----------|-----|-------|-----|---|
| | Itob ubtilebb | I Courto | ••• | I uuuuuu | ~ , | ILL . | | - |

Assay

Assay of FSD and TDF in Entadfi capsules was performed by the proposed method and the % assay of the both drugs were calculated as an average of 3 determinations, results were given in Table 10. These results indicate that the present HPLC method can be successfully used for the simultaneous assay of FSD and TDF in bulk and dosage forms. The data is tabulated in **Table 9**

 Table 9: Results of the market product

| Finasteride | Amount | <mark>% Assay ±</mark> | Tadalafil | Amount | <mark>% Assay ±</mark> |
|---------------------|-------------------------|------------------------|---------------------|-------------------------|------------------------|
| Label claim (mg) | found(mg) ± SD (n=3) | SD (n=3) | Label claim (mg) | found(mg) ± SD (n=3) | SD (n=3) |

| 5 | $\underline{4.99\pm0.08}$ | 99.85 ± 1.28 | 5 | 4.97 ± 0.14 | 99.42 ± 0.65 |
|---|---------------------------|-------------------|---|-----------------|------------------|
| 5 | 4.97 ± 0.05 | 98.64 ± 0.73 | 5 | 5.03 ± 0.27 | 100.2 ± 0.27 |
| 5 | 5.04 ± 0.10 | 100.82 ± 1.16 | 5 | 4.96 ± 0.35 | 99.24 ± 1.25 |

Forced Degradation Studies

The ability of the proposed method to separate FSD and TDF from their degradation products was evaluated by intentional degradation via various stress conditions such as acid hydrolysis (using 1N HCl), base hydrolysis (using 1N NaOH), oxidation (using 3% V/V H2 O2) at 60°C, thermal degradation at 110°C and data was given in **Table 10** and Shown in Figures **6,7,8,9 & 10**.

Preparation of stock solution

Accurately weigh and transfer 5mg of Finasteride, 5mg of Tadalafil working standard into a 10 ml clean dry volumetric flask add diluent and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Acid degradation

Pipette 1 ml of above solution into a 10ml volumetric flask and 1 ml of 1N Hcl was added. Then, the volumetric flask was kept at 60°C for 6 hours and then neutralized with 1 N NaOH and make up to 10ml with diluent. Filter the solution with 0.45 microns syringe filters and place in vials.



Figure 6: Chromatogram of Acid degradation

Alkali degradation

Pipette 1 ml of above solution into a 10ml volumetric flask and add 1ml of 1N NaOH was added. Then, the volumetric flask was kept at 60°C for 6 hours and then neutralized with 1N Hcl and make up to 10ml with diluent. Filter the solution with 0.45 microns syringe filters and place in vials.



Figure 7: Chromatogram of Alkaline degradation

Peroxide degradation

Pipette 1 ml above stock solution into a 10ml volumetric flask, 1 ml of 3% v/v of hydrogen peroxide added in 10 ml of volumetric flask and the volume was made up to the mark with diluent. The volumetric flask was then kept at room temperature for 15 min. Filter the solution with 0.45 microns syringe filters and place in vials.



Figure 8 : Chromatogram of Peroxide degradation

Thermal induced degradation

Finasteride and Tadalafil samples were taken in Petridish and kept in Hot air oven at 110° C for 24hours. Then the sample was taken and diluted with diluents and injected into HPLC and analysed.



Figure 9 : Chromatogram of Thermal degradation

Hydrolysis degradation

Pipette 1ml above stock solution into a 10ml volumetric flask, 1 ml of water added in 10 ml of volumetric flask and the volume was made up to the mark with diluent. The volumetric flask was then kept at room temperature for 15 min. Filter the solution with 0.45 microns syringe filters and place in vials



Figure 10: Chromatogram of Hydrolysis degradation

| | Fina | asteride | Tadalafil | | |
|--------------------------------------|---------|------------------|-----------|---------------|--|
| Results: % Degradation results | Area | % Degradation | Area | % Degradation | |
| Control | 2365530 | 0 | 2326806 | 0 | |
| Acid | 2034763 | 14.0 | 2033670 | 12.6 | |
| Alkali | 2052274 | 13.2 | 2017863 | 13.3 | |
| Peroxide | 1979715 | 16.3 | 1959113 | 15.8 | |
| Thermal | 2123289 | 10.2 | 2061184 | 11.4 | |
| Hydrolysis | 2306377 | 2.5 | 2301412 | 1.1 | |

| | - | | | _ | - | | | | | |
|--------|-----|----------|-------------|----------|-----|-----------|--------|-------|--------|----------|
| Tabla | Λ. | Formand | Dogwodotion | MOGINIEG | for | Finacto | wide e | and ' | Toda | ofil |
| гяше | 91 | rorcea | пергянянов | resillis | IOF | пняме | rice > | 41141 | гяня | X |
| I GOIC | · • | I UI CCU | Degradation | I Coulto | 101 | I III MOU | liuc i | | I HUHH | CCTTT |

CONCLUSION

The validated RP-HPLC-PDA method developed for the quantitative estimation of FSD and TDF in combination was evaluated for system suitability, specificity, sensitivity, linearity, range, accuracy (recovery), precision (repeatability and intermediate precision), and robustness. All the validation results were within the specifications of the ICH guidelines. The developed method has proven to be rapid, accurate, and stability-indicating for the simultaneous determination of the combined FSD and TDF in the capsule dosage form in the presence of excipients and the degradation products and always a complete separation of both ingredients from their

degradation products and from the placebo. Therefore, this method can be employed in quality control to estimate the amount of FSD and TDF in bulk and in combined dosage forms.

Abbreviations

FSD: Finasteride

TDF: Tadalafil

Conflicts of interest

There are no conflicts to declare

ACKNOWLEDGEMENTS

The authors are thankful to Rakshith pharmaceuticals and Dr.Reddy's Laboratories, Hyderabad for providing gift samples of FSD and TDF and also to the NRI College of Pharmacy, Pothavarappadu for providing facilities to carry out the research work

REFERENCES

- Ramakrishna NV, Vishwottam KN, Puran S. Quantification of Tadalafil in human plasma by liquid chromatography tandem mass spectrometry with electro spray ionization. J Chromatogr B Analyt Technol BiomedLife Sci. 2004;809:243–249.
- Rabbaa-Khabbaz L, Daoud RA. A Sensitive and Simple High Performance Liquid Chromatographic Method for Quantification of Tadalafil in Human Serum. *J Appl Res.* 2006;6:170–175.
- Gao W, Zhang Z, Li Z, Liang G. Chiral Separation of Two Pairs of Enantiomers of Tadalafil by High Performance Liquid Chromatography. *J Chromatogr Sci.* 2007;45:540– 543.
- Farthing CA, Farthing DE, Koka S, Larus T, Fakhry I, Xi L, Kukreja RC, Sica D, Gehr TW. A simple and sensitive HPLC fluorescence method for determination of Tadalafil in mouse plasma. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2010;878:2891–2895.

- Barot TG, Patel PK. Determination of Tadalafil in pure powder and tablet dosage form by high-performance liquid chromatography. *J AOAC Int.* **2010**;93:516–522.
- Mehanna MM, Motawaa AM, Samaha MW. Quantitation of transdermal Tadalafil in human skin by reversed-phase high-performance liquid chromatography. *J AOAC Int.* 2012;95:1064–1068.
- T.Takano and S. Hata, "High-Performance Liquid Chromatographic Determination of Finasteride in Human Plasma Using Direct Injection with Column Switching," Journal of Chromatography B: Biomedical Sciences and Applications, Vol. 676, No. 1, 1996, pp. 141-146.
- P. Ptácek, J. Macek and J. Klíma, "Determination of Finasteride in Human Plasma by Liquid-Liquid Extraction and High-Performance Liquid Chromatography," Journal of Chromatography B: Biomedical Sciences and Applications, Vol. 738, No. 2, 2000, pp. 305-310
- G. Carlucci and P. Mazzeo, "Finasteride in Biological Fluids: Extraction and Separation by a Graphitized Carbon Black Cartridge and Quantification by High-Performance Liquid Chromatography," Journal of Chromatography B: Biomedical Sciences and Applications, Vol. 693, No. 1, 1997, pp. 245-248.
- X. Chen, E. R. Gardner, D. K. Price and W. D. Figg, "Development and Validation of an LC-MS Assay for Finasteride and Its Application to Prostate Cancer Prevention Trial Sample Analysis," Journal of Chromatographic Science, Vol. 46, No. 4, 2008, pp. 356-361.
- Almeida, S. Almeida, A. Filipe, S. Gagnon, A. Mirapeix, B. Girard and M. Tanguay, "Bioequivalence Study of Two Different Coated Tablet Formulations of Finasteride in Healthy Volunteers," Arzneimittel-Forschung, Vol. 55, No. 4, 2005, pp. 218-222.
- F. Q. Guo, L. F. Huang, K. P. Wong, Y. H. Dai, Y. W. Li, Y. Z. Liang, K. L. Huang, K. J. Zhong and M. J. Wu, "A Rapid, Simple, Specific Liquid Chromatographic-Electrospray Mass Spectrometry Method for the Determination of Finasteride in Human Plasma and Its Application to Pharmacokinetic Study," Journal of Pharmacy and Biomedical Analysis, Vol. 43, No. 4, 2007, pp. 1507-1513.

- M. L. Constanzer, C. M. Chavez and B. K. Matuszewski, "Picogram Determination of Finasteride in Human Plasma and Semen by High-Performance Liquid Chromatography with Atmospheric-Pressure Chemical-Ionization Tandem Mass Spectrometry," Journal of Chromatography B: Biomedical Applications, Vol. 658, No. 2, 1994, pp. 281-287.
- ICH, "Stability Testing of New Drug Substances and Products (Q1AR)," International Conference on Harmonization, IFPMA, Geneva, 2000.
- ICH Draft Guidelines on Validation of Analytical Procedures: Definitions and Terminology," Federal Register, Vol. 60, No. 40, **1995**, p. 11260.

| S.No | Finasteride | | Tadalafil | |
|----------------------------|---------------|------------------|---------------------|-----------|
| | Conc.(µg/ml) | Pea k area | Conc.(µg/ml) | Peak area |
| 1 | 12.50 | 588487 | 12.50 | 618837 |
| 2 | 25.00 | 1161250 | 25.00 | 1222790 |
| 3 | 37.50 | 1751664 | 37.50 | 1718985 |
| 4 | 50.00 | 2330781 | 50.00 | 2361081 |
| 5 | 62.50 | 2863848 | 62.50 | 2952775 |
| 6 | 75.00 | 3519293 | 75.00 | 3504261 |
| Regressio n equation | y = 46494 x - | +1793 | y = 46307 x + 37204 | |
| Slope | 4649 | 4 | 46307 | |
| Intercept | 1793 | | 37204 | |
| R ² | 0.99 | 9 | 0.999 | |

Table 1: Linearity of Finasteride and Tadalafil

| S. No. | Area for Finasteride | Area for Tadalafil |
|-----------------------|----------------------|--------------------|
| 1 | 2359715 | 2341128 |
| 2 | 2380789 | 2352087 |
| 3 | 2369223 | 2344278 |
| 4 | 2345136 | 2365687 |
| 5 | 2389054 | 2311713 |
| 6 | 2351732 | 2332174 |
| Average | 2365942 | 2341178 |
| Standard Deviation | 16964.39 | 18322.49 |
| %RSD | 0.72 | 0.78 |

| Table 2: Method | l Precision | for Finasteride | & Tadalafil |
|-----------------|-------------|-----------------|-------------|
|-----------------|-------------|-----------------|-------------|

Table 3: Accuracy results of Finasteride

| %Concentration (at specification Level) | Area | Amount Added (mg) | Amount Found (mg) | % Recovery | Mean Recovery |
|---|---------|-------------------------|-------------------------|---------------|------------------|
| 50% | 1183115 | 2.5 | 2.5 | 100.0 | |
| 100% | 2355687 | 5 | 4.98 | 99.6 | 99.57 |
| 150% | 3515124 | 7.5 | 7.43 | 99.1 | |

Table 4: The Accuracy results for Tadalafil

| %Concentratio n (at specification Level) | Area | Amount Added (mg) | Amount Found (mg) | % Recovery | Mean Recovery |
|---|---------|-------------------------|-------------------------|---------------|------------------|
| 50% | 1180354 | 2.5 | 2.53 | 101.2 | |
| 100% | 2328120 | 5 | 5 | 100.0 | 100.1 |
| 150% | 3468324 | 7.5 | 7.44 | 99.2 | |

 Table 5: Sensitivity parameters (LOD & LOQ)
 Description

| Name of drug | LOD(µg/ml) | LOQ(µg/ml) |
|-----------------|------------|----------------|
| Finasteride | 0.143 | 0.157 |
| Tadalafil | 0.472 | 0.534 |

 Table 6: System suitability parameters for Finasteride & Tadalafil

| S.No | Parameter | Finasteride | Tadalafil | Accepted limit |
|------|--------------------|-------------|-----------|------------------------|
| 1 | Capacity factor(K) | 3.67 | 2.64 | <mark>></mark> 2 |
| 2 | Plate count(N) | 5349 | 5876 | <mark>>2000</mark> |
| 3 | Tailing factor(Tf) | 1.15 | 1.04 | <mark>< 2.0</mark> |
| 4 | Resolution(Rs) | | 21.50 | <mark>>2.0</mark> |
| 5 | %RSD | 0.52 | 0.48 | <mark>< 2.0%</mark> |

| | Finasteride | | | | | | | |
|----------------------------|----------------------|------------------------|-----------|------------|-------------|----------------|--|--|
| Parameter | Condition | Retention time(min) | Peak area | Resolution | Tailin g | Plate count | | |
| Flow rate | Less flow (0.8ml) | 2.684 | 2553564 | | 1.19 | 5444 | | |
| Change | Actual (1ml) | 2.415 | 2359707 | | 1.17 | 5342 | | |
| (mL/min) | More flow (1.2ml) | 2.290 | 2194138 | | 1.12 | 5279 | | |
| - · · | Less Org (63:37) | 2.700 | 2756501 | | 1.16 | 5470 | | |
| Organic Phase change | Actual (70:30) | 2.418 | 2380724 | | 1.15 | 5355 | | |
| | More Org (77:23) | 2.225 | 1965248 | | 1.08 | 5247 | | |

Table7:Robustness results of Finasteride by RP-HPLC

Table 8: Robustness results of Tadalafil by RP-HPLC

| D | Tadalafil | | | | | | | |
|-----------|----------------------|------------------------|-----------|------------|---------|----------------|--|--|
| Parameter | Condition | Retention time(min) | Peak area | Resolution | Tailing | Plate count | | |
| Flow rate | Less flow (0.8ml) | 7.685 | 2544562 | 23.01 | 1.13 | 5972 | | |
| Change | Actual (1ml) | 7.263 | 2325171 | 21.53 | 1.09 | 5869 | | |
| (mL/min) | More flow (1.2ml) | 6.958 | 2073124 | 22.68 | 1.06 | 5808 | | |
| Organic | Less Org | 7.711 | 2627541 | 23.18 | 1.14 | 5942 | | |

| Phase change | (63:37) | | | | | |
|--------------|---------------------|-------|---------|-------|------|------|
| | Actual (70:30) | 7.269 | 2339721 | 21.50 | 1.10 | 5862 |
| | More Org (77:23) | 6.606 | 1822874 | 21.30 | 1.04 | 5776 |

 Table 9: Forced Degradation results for Finasteride and Tadalafil

| | Fin | asteride | Tadalafil | | |
|--------------------------------------|---------|------------------|-----------|---------------|--|
| Results: % Degradation results | Area | % Degradation | Area | % Degradation | |
| Control | 2365530 | 0 | 2326806 | 0 | |
| Acid | 2034763 | 14.0 | 2033670 | 12.6 | |
| Alkali | 2052274 | 13.2 | 2017863 | 13.3 | |
| Peroxide | 1979715 | 16.3 | 1959113 | 15.8 | |
| Thermal | 2123289 | 10.2 | 2061184 | 11.4 | |
| Hydrolysis | 2306377 | 2.5 | 2301412 | 1.1 | |

 Table 10: Results of the market product

| Finasteride | Amount | <mark>% Assay ±</mark> | Tadalafil | Amount | <mark>% Assay ±</mark> |
|---------------------|--------------------------|------------------------|-------------|--------------------------|------------------------|
| Label claim (mg) | <mark>found(mg) ±</mark> | SD (n=3) | Label claim | <mark>found(mg) ±</mark> | SD (n=3) |
| | <mark>SD (n=3)</mark> | | (mg) | <mark>SD (n=3)</mark> | |
| 5 | $\frac{4.99\pm0.08}{}$ | 99.85 ± 1.28 | 5 | 4.97±0.14 | 99.42 ± 0.65 |
| 5 | 4.97 ± 0.05 | 98.64 ± 0.73 | 5 | 5.03 ± 0.27 | 100.2 ± 0.27 |
| 5 | 5.04 ± 0.10 | 100.82 ± 1.16 | 5 | 4.96 ± 0.35 | 99.24 ± 1.25 |